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Notes

NOVEL MACROLIDES FROM MICROMONOSPORA ROSARIA

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Rosaramicin (3), a sixteen-membered macrolide antibiotic, produced by *Micromonospora rosaria*, has been the subject of several investigations^{1~7}. The antibiotic is highly active against Gram-positive and Gram-negative bacteria. During the course of large scale fermentation besides 3, several related macrolides were isolated and characterized. Some of these have been previously reported^{8~12}. However, the carboxylic acid macrolide, which was expected from a natural sequence of events (eq 1)

$$\begin{array}{c} \operatorname{RCH}_2\operatorname{CH}_3 \longrightarrow \operatorname{RCH}_2\operatorname{CH}_2\operatorname{OH} \longrightarrow \operatorname{RCH}_2\operatorname{CHO} \\ \longrightarrow \operatorname{RCH}_2\operatorname{COOH} \longrightarrow \operatorname{RCH}_3 \quad (\text{eq 1}) \end{array}$$

has never been reported. We report here the isolation of this acid and other minor co-fermentation products. In addition, we also report the ¹³C NMR data (Table 1) of all the macrolides (Fig. 1).

Results and Discussion

Physical constants are presented in Table 2. Compound 4 ($C_{31}H_{51}NO_{10}$, MW m/z 597), UV λ_{max}^{EtOH} at 238 nm (ε 13,120), $[\alpha]_D^{26}$ -24.9° (c 0.23 EtOH) was isolated from the fermentation broth using XAD-4 resin and subsequently purified *via* silica gel column chromatography. ¹³C NMR (CDCl₃, ppm) indicated the presence of the methyl carbons at δ 8.6, 9.0, 14.5, 14.8 and 17.6, the methylene carbons at δ 24.5, 32.0, 35.9 and 40.1,

Table 1. ¹³C NMR data of rosaramicins in CDCl₃ (δ , ppm).

Carbon	1	1a	1b	2	3 ª	3a	3b	4	5	6	6a	7	7a	8	9	10	11
1	174.0	174.6	174.0	173.5	173.5	174.1	173.6	172.7	174.1	175.5	175.3	173.6	169.8	174.1	165.5	165.5	174.5
2	38.6	39.8	38.7	39.8	39.7	39.6	39.6	40.1	39.7	41.1	41.0	39.7	38.2	39.6	120.7	120.4	40.2
3	67.1	67.7	67.2	67.1	66.8	67.2	66.7	66.9	67.2	67.4	67.2	65.7	68.4	67.0	151.0	150.9	67.3
4	41.0	40.5	41.0	41.1	41.2	41.0	41.1	42.8	41.8	42.1	42.2	41.1	40.6	41.1	41.8	41.5	42.2
5	80.2	80.2	81.0	81.2	81.3	81.2	81.8	83.8	86.7	80.3	80.3	80.6	80.5	81.7	82.5	82.0	88.6
6	39.9	38.7	39.9	33.1	31.4	31.4	31.4	33.6	31.4	35.4	35.4	32.8	33.7	36.2	31.1	33.6	69.4
7	33.2	33.9	33.4	31.7	31.8	32.5	31.9	32.0	34.2	38.1	38.2	32.1	32.2	32.8	31.1	30.7	36.4
8	45.2	44.9	45.3	45.3	45.1	45.1	45.1	45.3	45.3	49.6	49.3	45.0	44.7	45.2	45.1	45.6	45.6
9	201.3	204.2	201.4	201.5	200.3	203.3	200.6	201.5	201.2	160.0	159.9	201.1	200.4	200.9	200.7	200.3	203.5
10	123.2	118.9	123.2	123.2	122.8	118.3	122.9	123.0	123.3	111.8	111.8	123.1	123.4	122.9	122.9	122.8	125.1
11	150.2	145.2	150.4	150.6	150.9	146.0	151.0	150.7	150.0	161.9	161.9	150.6	150.5	150.7	150.9	151.5	151.4
12	59.8	133.6	59.8	59.7	59.7	133.4	59.8	59.8	59.8	62.5	62.5	59.7	59.9	59.7	60.1	60.1	61.0
13	67.9	147.5	68.2	68.1	68.0	148.2	68.0	68.6	67.8	72.1	72.2	68.0	68.4	68.0	66.9	66.8	69.3
14	37.8	38.7	37.9	37.7	37.8	38.9	37.8	37.8	37.8	36.8	36.7	37.7	36.7	37.8	37.2	37.2	38.7
15	76.7	78.7	76.8	76.7	76.8	78.6	76.9	76.3	76.9	79.2	79.2	76.7	77.3	76.9	76.3	76.2	78.2
16	24.7	24.7	24.8	24.7	24.7	24.8	24.7	24.5	24.8	25.9	25.9	24.7	24.4	24.7	24.9	25.0	25.5
17	9.0	9.0	9.1	9.0	9.0	8.8	8.9	8.6	9.0	8.5	8.5	9.0	8.5	9.0	8.9	8.9	9.4
18	9.2	9.6	9.6	9.1	9.0	9.7	9.1	9.0	9.3	9.4	9.5	9.2	10.5	9.1	19.1	18.8	9.5
19	21.1	21.0	21.3	33.1	43.9	43.7	43.9	35.9	17.5	138.4	138.3	35.7	34.3	32.3	39.5	44.6	
20	12.0	12.0	12.0	60.7	202.9	203.5	203.2	180.3		144.6	144.6	67.2	67.6	149.0	148.0	201.8	
21	17.5	17.6	17.5	17.4	17.4	17.6	17.4	17.6	17.4	18.3	18.2	17.4	17.6	17.4	17.4	17.4	17.7
22	15.0	16.2	15.0	14.9	15.0	16.2	15.0	14.8	15.0	16.1	16.1	14.9	15.5	14.9	15.8	15.9	15.3
23	14.5	12.9	14.5	14.4	14.5	12.9	14.5	14.5	14.5	14.8	14.9	14.5	14.6	14.5	14.6	14.6	14.7
24												50.7	48.3	132.3	132.9		
25												209.2	206.3	199.0	198.8		
26												31.1	30.8	26.6	27.1		

^a Assignments based upon ¹³C-¹³C INADEQUATE data.

Fig. 1. Rosaramicin macrolides.



- 1 $R = CH_3$; 1a 12,13-desepoxy; 1b N-oxide
- $2 R = CH_2OH$
- 3 R = CHO; 3a 12,13-desepoxy; 3b N-oxide
- $4 \quad R = COOH$
- 5 R = H
- 6 "2N" component; 6a N-oxide
- 7 $R = -CH(OH)CH_2COCH_3$; 7a 3,20,2'-triacetate
- 8 $R = -CH = CHCOCH_3$
- 9 R=-CH=CH-CO- CH_3 , Δ^2
- 10 R = CHO, Δ^2
- 11 C-6=OH; 11a 3,6,2'-triacetate

the methine carbons at δ 33.6, 37.8, 42.8, 45.3, 66.9, 68.6, 76.3 and 83.8 and the olefinic and singlet carbons at δ 59.8, 123.0, 150.7, 172.7, 180.3 and 201.5. The comparison of the data with that of **3** indicated that C-20 has been oxidized from – CHO \rightarrow COOH (δ 202.9 \rightarrow 180.3) with concurrent perturbations in the chemical shifts of other carbons, *e.g.*, C-19 (δ 43.9 \rightarrow 35.9). ¹H NMR ((CD₃)₂CO, ppm) showed the presence of diagnostic resonances at δ 0.90 (t, CH₃), 1.10 (d, 2 × CH₃), 1.12 (d, CH₃), 1.20 (d, CH₃), 1.24 (d, CH₃), 1.50 (s, CH₃), 3.69 (m, – CHO–), 4.86 (m, – CHOCO), 6.43 (d, =CH) and 6.88 (d, =CH). The spectrum lacked the presence of a –CHO function (δ 9.76 in 3).[†]

HPLC of the bulk material led to the detection of a new co-fermentation product, purified by silica gel column chromatography (CH₂Cl₂-EtOH-H₂O-HOAc, 24:8:1:1). ¹³C NMR (CD₃OD-CDCl₃, ppm) of the purified product (**11**, MW *m/z* 555) indicated the presence of the methyl carbons at δ 9.4, 9.5, 14.7, 15.3 and 17.7, the methylene carbons at δ 25.5, 36.4 and 40.2, the methine carbons at δ 38.7, 42.2, 45.5, 67.3, 69.3, 69.4, 78.2 and 88.2 and the olefinic and singlet carbons at δ 61.0, 125.1, 151.4, 174.5 and 203.5. ¹H NMR ((CD₃)₂CO, ppm) indicated all the diagnostic resonances at δ 0.91 (t, CH₃), 1.15 (d, 2×CH₃), 1.22 (d, CH₃), 1.24 (d, CH₃) 1.50 (s, CH₃), 2.84 (d, -CHO), 4.92 (m, CHOCO), 6.40 (d, =CH) and 6.90 (d, =CH).

Comparison of the data of 5, 3 and 11 (Table 3) showed that in 5 and 11 C-5 and C-7 are shifted downfield compared to 3 where they are influenced by the γ -effect of C-20. The data are consistent with structure 11 (6-hydroxy) for this isolate. Treatment with acetic anhydride in pyridine resulted in the formation of 3, 6, 2'-triacetate (m/z 681.4) supported by ¹³C and ¹H data.

Spectroscopic Data

¹³C NMR spectrum of **5** (R=H) indicated the presence of a lactone group at δ 174.1, α,β unsaturated keto moiety at δ 201.2, 123.3 and 150.0, an epoxide ring at δ 59.8 and 67.8, three oxygen bearing methine carbons at δ 67.2 (HCOH), 76.9 (HCOCO) and 86.7 (HC-O-sugar), the latter having desosamine β -glycosidically linked. Desosamine ring does not undergo severe conformational changes and is discussed only in the case of

Table	2.	Physical	constants.
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	1	a	1	b	3	sb		4	6	a
Formula MW	C ₃₁ H 552 ()	$_{53}NO_{7}$ M + 1)	C ₃₁ H	53NO9 83	C ₃₁ H ₅	₅₁ NO ₁₀ 97	C ₃₁ H ₅	97	C ₃₁ H ₄ 5	_{.8} N ₂ O ₈ 76
Elemental	Calcd	Found	Calcd	Found	Calcd	Found	Calcd	Found	Calcd	Found
Anal										
С	67.48	66.62	63.78	62.37	62.29	57.73	62.29	59.60	64.56	60.07
н	9.68	9.27	9.15	8.59	8.60	8.05	8.60	7.82	8.39	8.35
Ν	2.54	2.64	2.40	2.45	2.34	2.21	2.34	2.56	4.86	4.23
UV $\lambda_{max}^{E(OH)}$ nm	2	82	238	, 283	2	38	2	38	238, 2	78, 287
8	20,	400	4,750	, 1,540	12	,130	13,	,120	7,100, 2,	050, 1,780
$[\alpha]_{D}^{26}$ (c 0.2~0.5 EtOH)	- 1	1.8°	-2	2.3°	. — 3	9.7°	-2	.4.9°	-	_

[†] The ¹³C and ¹H NMR data for desosamine sugar were not affected and are not discussed unless otherwise specified.

Carbon	Compounds							
Carbon	5	3	11					
4	41.8	41.3	42.2					
5	86.7	80.9	88.6					
6	31.4	31.7	69.4					
7	34.2	31.7	36.4					
8	45.3	45.5	45.6					
9	201.2	201.7	203.7					

Table 3^a. ¹³C NMR chemical shifts of 5, 3 and 11.

Chemical shifts given in ppm.

^a Solvent: CDCl₃ (5); CD₃OD - CDCl₃ (3 and 11).

N-oxidation. The resonances at δ 31.4 and 17.5 are attributed to carbons 6 and 19, respectively.

For RCH₃, three compounds 1 (R=CH₃), 1a (12,13-desepoxy) and 1b (desosamine *N*-oxide) were isolated. The introduction of a methyl group (δ 12.0) at C-19 shifted C-6 to 39.9 ($\Delta \delta$ = +8.5 ppm) and C-19 to 21.1 ($\Delta \delta$ = +3.6 ppm). C-5 suffered an upfield shift ($\Delta \delta$ = -6.5 ppm), a fact attributed to a combination of γ gauche effect and steric compression.

Desepoxidation in 1a resulted in an additional double bond leading to a five carbon conjugation and significant planarity in one-half of the aglycone. Methyl-23 was shifted upfield ($\Delta \delta = -1.6 \text{ ppm}$) as expected. Compound 1b was more polar than 1 and the ¹³C NMR analysis indicated *N*-oxidation (discussed separately). The presence of a polar group (R=CH₂OH) in 2, shifted C-19 to 33.1 vs. 21.1 in 1 and the CH₂OH carbon appeared at δ 60.7. C-6 experienced an upfield shift to 33.1 from 39.9 in 1.

Further oxidation resulted in rosaramicin 3 (R=CHO), the major and the most important isolate of the fermentation. C-19 shifted further downfield from $21.2 \rightarrow 43.9$ with concurrent generation of a CHO function at $\delta 202.9$ ppm. Desepoxy rosaramicin (3a) has aglycone carbon shifts similar to 1a and the *N*-oxide (3b) has chemical shifts of desosamine identical to that of the same in 1b. C-6 appeared upfield at $\delta 31.4$ in 3, 3a and 3b compared to $\delta 39.9$ ppm in 1, 1a and 1b. Compound 4 (R=COOH) has never been mentioned before because of the difficulty in its isolation. The acid was isolated from the aqueous mother liquor by precipitation. C-19 and C-20 appeared at $\delta 35.9$ and 180.3, respectively.

The structure of "2N" impurity **6** has been reported earlier⁸⁾. However, further improvement in the methodology led to the isolation of N-oxide (**6a**).



Compound 7 isolated as an acetone adduct, in which 20-CHO function underwent aldol reaction:



In 7, new carbons appeared at δ 67.2 (CHOH), 209.2 (C=O), 50.7 (CH₂) and 31.1 (CH₃). Further characterization was achieved by converting 7 to its 3,20,2'-triacetate (7a) with expected chemical shift changes except for an upfield shift of C=O (209.2 \rightarrow 206.3) and ester (COO) (173.6 \rightarrow 169.8) carbons, indicative of the loss of intramolecular hydrogen bonding.

¹³C T₁ relaxation data for 7 was obtained for comparison with that of $3^{4)}$. The average NT₁ values for the aglycone and desosamine moieties were 0.50 ± 0.21 and 0.51 ± 0.11 seconds, respectively. The same values for 3 were 0.40 ± 0.05 seconds and 0.41 ± 0.03 seconds, respectively. Compounds 7 and 3 undergo isotropic globular motion in solution. However, the side chain at C-6 displayed increased mobility as shown by the following T₁ values.



High T_1 value (1.15 seconds) for methyl-22 suggested free rotation even though methyl groups at quarternary carbons in 14-membered macrolides were restricted in their mobility.

Dehydration of 7 led to the isolation of another UV absorbing component which was assigned structure, 8, based upon the presence of two new olefinic carbons at δ 149.0 (C-20) and 132.3 (C-24) and shifting COCH₃ carbons to 199.0 and 26.6 ppm,



respectively. Attempts to prepare 8 from 7 via triacetate, 7a, followed by dehydration led to the isolation of 9. Compound 9 had three α,β unsaturated keto moieties encompassing C-1, C-2, C-3, C-9, C-10, C-11 and C-20, C-24, and C-25. The double bond at 2,3-position (Δ^2) resulted in the appearance of olefinic carbons at δ 120.7 and 151.0 with concurrent upfield shift of the ester carbonyl (δ 174.4 \rightarrow 165.5; loss of intramolecular hydrogen bond and unsaturation effect). Except for the presence of C-20, C-24 double bond and its proximity effects, the data of 9 were comparable to that of Δ^2 -rosaramicin (10).

Incorporation (13 C- and 14 C-labeled precursors) studies indicated that carbons 20, 19, 6 and 5 of the aglycone moiety of **3** originated from a butyrate molecule⁶⁾ and the isolation of all components in the sequence,



suggests successive oxidation during fermentation. Similar transformations have been observed for rifamycins antibiotics¹³⁾. Attempts to establish the kinetics of this oxidation in **3** utilizing 4-¹³Clabeled butyrate were not successful.

Sugar Moiety

¹³C Chemical shifts of desosamine are identical to the reported values and are consistent with all *trans* stereochemistry. However, *N*-oxidation imparts significant changes as a result of $N\rightarrow O$ bond orientation. The chemical shifts (δ , ppm) of the sugar carbons are shifted from, *e.g.*, 1' (104.5 \rightarrow 103.2), 2' (70.4 \rightarrow 72.2), 3' (65.8 \rightarrow 76.1), 4' (28.4 \rightarrow 34.9), 5' (69.7 \rightarrow 67.5), 6' (21.1 \rightarrow 21.0) and 7', 8' (40.2 \rightarrow 52.2, 59.1). The influence of N–O bond for carbons 2', 3', 4', 7' and 8' suggests its usefulness as a diagnostic probe.

The *in vitro* antibacterial activity of **4** was $2 \sim 64$, and $> 64 \,\mu$ g/mg (MIC) for Gram-positive and Gram-negative organisms, respectively.

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